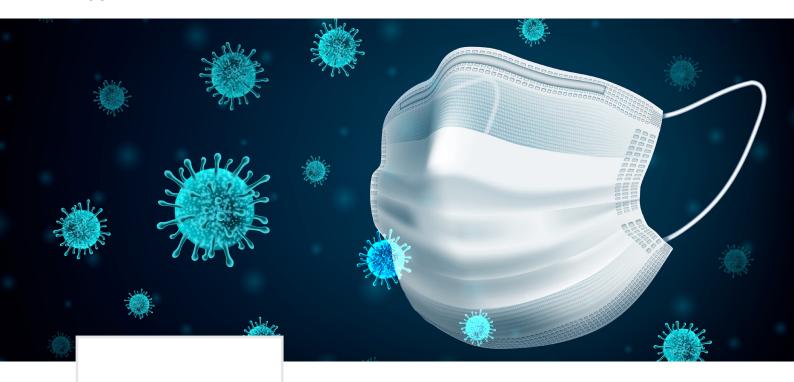
Application Note · UVP Crosslinker

Epak Electronics Ltd www.epakelectronics.com





Challenge

Access to a steady supply of N95 respirator masks is inadequate due to the COVID-19 pandemic

Solution

Irradiation and reuse of existing N95 masks using the UVP Crosslinker

Ultraviolet-based Disinfection of N95 Respirator Masks

Abstract

Amid the ongoing SARS-CoV-2 outbreak, despite efforts by the private sector and governments globally, access to a steady supply of N95 respirator masks is lacking. An important mitigation approach to the limited supply of N95 masks is the irradiation and reuse of existing N95 masks, which the CDC contends is a promising strategy, especially during a pandemic¹. Recently^{2,3}, a hospital-approved ultraviolet germicidal irradiation (UVGI) workflow for masks has become available. Ideally, healthcare workers (HCWs) should not routinely resort to reusing masks. However, if needed, the integration of the UVC-based CL-3000 crosslinker into a hospital-approved UVGI workflow will generate 1J/cm² of UVC with reproducible results.

Background

Shortwave ultraviolet light (UVC) has germicidal properties by acting directly on the DNA/RNA of microorganisms. DNA/RNA absorb ultraviolet light maximally at approximately 260 nm, which, as a consequence, damages the DNA/RNA structure. Although microorganisms have mechanisms with which to counteract this damage, they cannot overcome extensive high-intensity UV doses, which ultimately inactivate or kill the microorganism. Ultraviolet germicidal irradiation (UVGI) of viruses has been demonstrated extensively in the scientific literature for a range of viruses^{4-37‡}, and most recently for virus contaminated N95 filtering facepiece respirators (FFRs)^{2,3,7,27,34‡‡}.



To prepare for inevitable shortages of N95 FFRs during the ongoing SARS-CoV-2 pandemic, the University of Nebraska Medical Center (UNMC) and Stanford University have developed UVGI workflows to be implemented by HCWs^{2,3}. These workflows take advantage of UV equipment sometimes already found in the healthcare environment including, but not limited to, UV disinfection boxes³ and whole-room UV emitters^{2,15}. Some major concerns with these instruments are the lack of uniformity in germicidal light, the risk of inconsistent dosing between disinfection cycles, as well as the risk of UV exposure to HCWs in general^{38,39}. At Analytik Jena we believe we have a better solution.

Since 1993 we have been supplying the UVP Crosslinker (CL-1000/CX-2000) primarily for molecular biology applications, which are reliant on high-intensity, reproducible UVC doses. By having reflective housing and the UV source in close proximity to the sample (Figure 1A), we are able to achieve highly uniform illumination (Figure 1B and 1C). Our most recent model, the CL-3000 (Figure 1A), is designed with a built-in radiometer calibrated to a NIST traceable standard—this ensures consistent doses irrespective of space and time. In addition, the CL-3000 can produce a cumulative dose of up to 10J/cm². As with all our instruments, safety of the end user is critical, and all our Crosslinkers have a safety-interlock to prevent accidental UV exposure. Most importantly, there is an extensive body of scientific work where our Crosslinker is used for viral inactivation of viruses from several families^{11,18,19,24–26,31–33,35,36}, including coronaviruses^{21,23,30}, which is lacking for other instruments on the market.

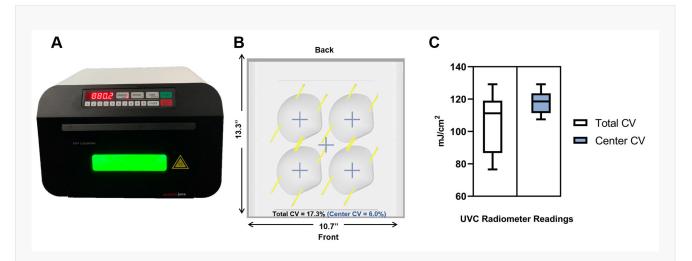


Figure 1. Uniformity of the Analytik Jena UVP Crosslinker equipped with 254 nm UVC bulbs. A) Profile image of the UVP CL-3000 Crosslinker during disinfection cycle. B) Diagram depicting position of integrated radiometer readings in mJ/cm² with dimensions and calculated coefficient of variation (CV) for the entire illumination surface (Total CV), and the center of the illumination surface (Center CV), the latter of which is where masks should be placed. C) Box-whisker plot summary of radiometer readings. Total CV: Mean =104.9, SD= 18.16; Center CV: Mean = 117.9, SD=7.1. We recommend users place masks in the center of the illumination to maximize evenness of UV exposure. Samples place in the center will receive, at a minimum, the input energy dose. If users want to place samples at the front-most and back-most positions, we recommend purchasing a UVP 254 nm radiometer (P/N 97-0015-02) and sensor (P/N 97-0016-01) to calibrate dose.

At Analytik Jena, we agree with the CDC, which contends that UVGI is a viable crisis strategy for capacity management of personal protective equipment¹. In summary, our Crosslinker can be easily integrated into UVGI mask disinfection workflows for the following reasons:

- Fixed distance between sample and UVC source to ensure uniform dosing
- Embedded radiometer calibrated to NIST traceable standards to ensure reproducible dosing
- Higher UVC output for shorter disinfection cycles
- Safety Interlock to prevent accidental exposure
- Small footprint to accommodate limited spaces and/or decentralization
- Closed system to accommodate scaling up strategies
- Proven history of performance from the scientific literature

Operating the CL-3000 as part of an approved UVGI Workflow

According to an FDA commissioned report, a dose of $1J/cm^2$ is sufficient to result in no detectable virus on N95 mask material after UVGI treatment, corresponding to a 3.9-4.5 log₁₀ reduction for MERS-CoV and 4.0-4.8 log₁₀ reduction for SARS-CoV, between experimental and control groups³⁴. Below, we simply describe how to use our device to achieve the same irradiation level of $1J/cm^2$. HCWs should inspect masks for wear and tear after each disinfection cycle^{2,3,13,34}.

To operate the CL-3000 simply follow the instructions below:

- 1. Set the dosage on the instrument by selecting ENERGY, then pressing 1-0-0-0 for 1000.0 mJ/cm² (or 1J/cm²), and then press ENTER.
- 2. Place mask into the center-most portion of the chamber and close door.
- 3. Press START.
- 4. Open door and flip mask over and repeat step 3.
- 5. Remove the sample and continue following the UVGI protocol approved by your hospital/institution.

Note: As an added precaution, users may consider repeating steps 1-3 with an empty crosslinker in between disinfection cycles if residual contamination is a concern.

Disclaimer: We do not advocate specific treatments or approaches. We are simply sharing the most recent evidence from the medical community to help HCWs during the SARS-CoV-2 pandemic. Your UVGI workflow should be set and approved by your hospital/institution. Users can refer to the UNMC² and/or Stanford University³ workflows for process recommendations.

Technical Data

Technical Specifications	CL-3000
Wavelength	254nm
Bulbs	6 x 8 Watt
Energy	0000.1 - 9999.9 mJ/cm² (0 - 10 J/cm²)
Time	000:01 - 999:59 mmm:ss (>300J/cm²)
Temperature	15°C - 35°C
Humidity	70% Non-Condensing
Altitude	up to 3,000M (9,842 ft)
Sound Level	≤ 50 dba
Housing Surface Temp	≤ 30°C
Startup Time	< 1 sec
External Dim (L x W x H)	41cm x 40cm x 26.5cm
Internal Dim (L x W x H)	35cm x 27cm x 16cm
Weight	6.8Кg: 15 lb
Operating Power	100 - 115VAC & 230VAC 50/60Hz
Certifications	CE, RoHS (CSA In Process)

Part Numbers and Description

Part Number	Description
849-95-0615-01	UVP Crosslinker (CL-3000), 254 nm, 100 – 115V
849-95-0615-02	UVP Crosslinker (CL-3000), 254 nm, 230V

Mid-Range Wavelength: 302 nm (CL-3000M) and Long-Range Wavelength: 365 nm (CL-3000L) UVP Crosslinkers are available to order, but are not applicable to this application note.

References

- 1. CDC. Coronavirus Disease 2019 (COVID-19) Decontamination and Reuse of filtering Facepiece Respirators. Centers for Disease Control and Prevention https://www.cdc.gov/coronavirus/2019-ncov/hcp/ppe-strategy/decontamination-reuse-respirators.html (2020).
- 2. CLowe, J. et al. N95 Filtering Facemask Respirator Ultraviolet Germicidal Irradiation (UVGI) Process for Decontamination and Reuse. https://www.nebraskamed.com/sites/default/files/documents/covid-19/n-95-decon-process.pdf (2020).
- 3. Price, A. & Chu, L. COVID-19 Evidence Service Addressing COVI-19 Face Mask Shortages [v1.2]. https://aim.stanford. edu/covid-19-evidence-service/ (2020).
- 4. Darnell, M. E. R., Subbarao, K., Feinstone, S. M. & Taylor, D. R. Inactivation of the coronavirus that induces severe acute respiratory syndrome, SARS-CoV. Journal of Virological Methods 121, 85–91 (2004).
- 5. McDevitt, J. J., Milton, D. K., Rudnick, S. N. & First, M. W. Inactivation of Poxviruses by Upper-Room UVC Light in a Simulated Hospital Room Environment. PLoS ONE 3, e3186 (2008).
- 6. Rutala, W. A., Gergen, M. F. & Weber, D. J. Room Decontamination with UV Radiation. Infect. Control Hosp. Epidemiol. 31, 1025–1029 (2010).
- 7. Fisher, E. M. & Shaffer, R. E. A method to determine the available UV-C dose for the decontamination of filtering facepiece respirators. Journal of Applied Microbiology 110, 287–295 (2011).
- 8. Park, G. w., Linden, K. g. & Sobsey, M. d. Inactivation of murine norovirus, feline calicivirus and echovirus 12 as surrogates for human norovirus (NoV) and coliphage (F+) MS2 by ultraviolet light (254 nm) and the effect of cell association on UV inactivation. Letters in Applied Microbiology 52, 162–167 (2011).
- 9. Zou, S. et al. Inactivation of the novel avian influenza A (H7N9) virus under physical conditions or chemical agents treatment. Virol J 10, 289 (2013).
- 10. Kiryu, I., Sakai, T., Kurita, J. & Iida, T. Virucidal Effect of Disinfectants on Spring Viremia of Carp Virus. Fish Pathol. 42, 111–113 (2007).
- 11. Singh, R., Sharma, A., Hong, S. & Jang, J. Electrical immunosensor based on dielectrophoretically-deposited carbon nanotubes for detection of influenza virus H1N1. Analyst 139, 5415–5421 (2014).
- 12. Beck, S. E., Wright, H. B., Hargy, T. M., Larason, T. C. & Linden, K. G. Action spectra for validation of pathogen disinfection in medium-pressure ultraviolet (UV) systems. Water Research 70, 27–37 (2015).
- 13. Lindsley, W. G. et al. Effects of Ultraviolet Germicidal Irradiation (UVGI) on N95 Respirator Filtration Performance and Structural Integrity. Journal of Occupational and Environmental Hygiene 12, 509–517 (2015).
- Bae, K. S., Shin, G.-A., Bae, K. S. & Shin, G.-A. Inactivation of various bacteriophages by different ultraviolet technologies: Development of a reliable virus indicator system for water reuse. Environmental Engineering Research 21, 350–354 (2016).
- 15. Bedell, K., Buchaklian, A. & Perlman, S. Efficacy of an automated multi-emitter whole room UV-C disinfection system against Coronaviruses MHV and MERS-CoV. Infect Control Hosp Epidemiol 37, 598–599 (2016).
- 16. Chu, H.-A. & Chiu, Y.-F. The Roles of Cytochrome b559 in Assembly and Photoprotection of Photosystem II Revealed by Site-Directed Mutagenesis Studies. Front. Plant Sci. 6, (2016).
- 17. Song, K., Mohseni, M. & Taghipour, F. Application of ultraviolet light-emitting diodes (UV-LEDs) for water disinfection: A review. Water Research 94, 341–349 (2016).
- 18. Ziegler, C. M. et al. The Lymphocytic Choriomeningitis Virus Matrix Protein PPXY Late Domain Drives the Production of Defective Interfering Particles. PLoS Pathog 12, (2016).
- 19. Berger, A. K. et al. Viral RNA at Two Stages of Reovirus Infection Is Required for the Induction of Necroptosis. J. Virol. 91, e02404-16, /jvi/91/6/e02404-16.atom (2017).

- 20. Fryk, J. J. et al. Reduction of Zika virus infectivity in platelet concentrates after treatment with ultraviolet C light and in plasma after treatment with methylene blue and visible light. Transfusion 57, 2677–2682 (2017).
- 21. Fung, T. S. & Liu, D. X. Activation of the c-Jun NH 2 -terminal kinase pathway by coronavirus infectious bronchitis virus promotes apoptosis independently of c-Jun. Cell Death & Disease 8, 1–13 (2017).
- 22. Kim, D.-K., Kim, S.-J. & Kang, D.-H. Inactivation modeling of human enteric virus surrogates, MS2, Qβ, and ΦX174, in water using UVC-LEDs, a novel disinfecting system. Food Research International 91, 115–123 (2017).
- 23. Bodmer, B. S., Fiedler, A. H., Hanauer, J. R. H., Prüfer, S. & Mühlebach, M. D. Live-attenuated bivalent measles virusderived vaccines targeting Middle East respiratory syndrome coronavirus induce robust and multifunctional T cell responses against both viruses in an appropriate mouse model. Virology 521, 99–107 (2018).
- 24. Frascaroli, G. et al. Human Macrophages Escape Inhibition of Major Histocompatibility Complex-Dependent Antigen Presentation by Cytomegalovirus and Drive Proliferation and Activation of Memory CD4+ and CD8+ T Cells. Front. Immunol. 9, (2018).
- 25. Lee, M. K. et al. Transmembrane Protein pUL50 of Human Cytomegalovirus Inhibits ISGylation by Downregulating UBE1L. J Virol 92, e00462-18, /jvi/92/15/e00462-18.atom (2018).
- 26. Mathew, A. M., Mun, A. B. & Balakrishnan, A. Ultraviolet Inactivation of Chikungunya Virus. Intervirology 61, 36–41 (2018).
- 27. Mills, D., Harnish, D. A., Lawrence, C., Sandoval-Powers, M. & Heimbuch, B. K. Ultraviolet germicidal irradiation of influenza-contaminated N95 filtering facepiece respirators. American Journal of Infection Control 46, e49–e55 (2018).
- Nishisaka-Nonaka, R. et al. Irradiation by ultraviolet light-emitting diodes inactivates influenza a viruses by inhibiting replication and transcription of viral RNA in host cells. Journal of Photochemistry and Photobiology B: Biology 189, 193–200 (2018).
- 29. Vaidya, V. et al. Ultraviolet-C irradiation for inactivation of viruses in foetal bovine serum. Vaccine 36, 4215–4221 (2018).
- 30. Yang, L. et al. Porcine Epidemic Diarrhea Virus-Induced Epidermal Growth Factor Receptor Activation Impairs the Antiviral Activity of Type I Interferon. J Virol 92, e02095-17, /jvi/92/8/e02095-17.atom (2018).
- 31. Baidaliuk, A. et al. Cell-Fusing Agent Virus Reduces Arbovirus Dissemination in Aedes aegypti Mosquitoes In Vivo. J Virol 93, e00705-19, /jvi/93/18/JVI.00705-19.atom (2019).
- 32. Campbell, T. M. et al. Functional paralysis of human natural killer cells by alphaherpesviruses. PLoS Pathog 15, (2019).
- 33. DeFord, D. M. et al. Evaluation of the role of respiratory syncytial virus surface glycoproteins F and G on viral stability and replication: implications for future vaccine design. Journal of General Virology, 100, 1112–1122 (2019).
- 34. Heimbuch, B. & Harnish, D. Research to mitigate a shortage of respiratory protection devices during public health emergencies. (2019).
- 35. Lee, A. C. Y. et al. H7N9 influenza A virus activation of necroptosis in human monocytes links innate and adaptive immune responses. Cell Death & Disease 10, 1–16 (2019).
- 36. Romero, N., Van Waesberghe, C. & Favoreel, H. W. Pseudorabies virus infection of epithelial cells leads to persistent but aberrant activation of the NF-κB pathway, inhibiting hallmark NF-κB-induced pro-inflammatory gene expression. J Virol JVI.00196-20, jvi;JVI.00196-20v1 (2020) doi:10.1128/JVI.00196-20.
- 37. Zhao, X. et al. Activation of C-Type Lectin Receptor and (RIG)-I-Like Receptors Contributes to Proinflammatory Response in Middle East Respiratory Syndrome Coronavirus-Infected Macrophages. J Infect Dis 221, 647–659 (2020).
- 38. Zaffina, S. et al. Accidental exposure to UV radiation produced by germicidal lamp: case report and risk assessment. Photochem. Photobiol. 88, 1001–1004 (2012).
- 39. International Commission on Non-Ionizing Radiation Protection. ICNIRP statement-Protection of workers against ultraviolet radiation. Health Phys 99, 66–87 (2010).

‡These citations do not represent the entire body of literature for UVGI of viruses and only serve to represent the diversity of viruses that have been inactivated with UVGI.

‡‡ As of 4/7/2020 Ref. 2 & 3 have only been published online and are not peer-reviewed.

This document is true and correct at the time of publication; the information within is subject to change. Other documents may supersede this document, including technical modifications and corrections.

Analytik Jena US LLC 2066 W. 11th Street Upland, CA 91786 · USA Phone +1 909 946 3197 Fax +1 909 946 3597 info@us.analytik-jena.com www.analytik-jena.com en · 04/2020 © Analytik Jena US LLC | Pictures ©: Analytik Jena / freepik Printout and further use permitted with reference to the source

Epak Electronics Ltd. Millfield Estate, Chard, Somerset TA20 2BB United Kingdom.www.epakelectronics.comsales@epakelectronics.comT: +44 01460 61791